

**INTERNATIONAL JOURNAL OF ADVANCES IN
PHARMACY, BIOLOGY AND CHEMISTRY**

Review Article

**Natural Nutritional Ingredients hold the key to a
Sustainable Future in Aquaculture: Awakening the
Indian Sundarbans for self preservation**

D. Chakravartty

Department of Oceanography, Techno India University, EM 4/1, Sector V,
Salt Lake, Kolkata, West Bengal, India - 700091.

Email: dechavv@gmail.com

ABSTRACT

With the ever expanding aquaculture sector, the demand for feed rises. This has led to the sprouting of a multitude of manufacturers, some even introducing spurious additives to make up for lack of protein content in feed and hence bridging a gap between feed formulators and aqua-farmers. The over exploitation of the aquaculture trade has adverse impacts on feed availability, disease and the ecology. The use of antibiotics have created further imbalance with drug resistant microorganisms. In an ongoing effort to reduce the use of crude feed and control of disease and drugs, FAO has advised the development of certain nutritional supplementations for a sustainable and profitable aquaculture process. As a response to this we have identified key natural nutritional components that may provide sustenance in aquaculture. Amino acids, allicin (from garlic) and sea weed extract have shown to provide a full spectrum of growth, immunity, antioxidant values and pose as alternatives to pharmaceutical antibiotics. We look into the prospects of these supplementments as a sustainable solution in aquaculture nutrition by reviewing the work done so far by investigators. The coastal aquaculture communities of the Indian Sundarbans need to adopt such options. The challenge is to build trust with aquafarmers for them to accept sustainability programmes with sound management action to envision a safer aquaculture domain with a greener Sundarbans along with a positive effort to sustain India's position in aquaculture production.

Key words: Sustainable aquaculture, amino acid, allicin, sea weed, Indian Sundarbans

INTRODUCTION

FAO predicts the global aquaculture production will exceed 100 million tonnes by 2020. With a current world production at 66 million tonnes ¹, India contributes over 4.2 million tonnes ranking them the world's second largest aquaculture producer. China sits at the top of the table at 41 million tonnes. With a continuous rising demand in fish and other aquaculture produce a reduction in operation costs and sustainability is of utmost importance.

With fast disappearing fresh water areas, aquaculture development gradually found its way into marine and brackish water systems ². Aquaculture of shrimp and

piscivorous fishes has several drawbacks. Intensive monoculture causes disaster to coastal ecosystems by way of deteriorating the water quality, mangrove deforestation etc ^{3, 4, 5}. As a result of over exploitation, GreenPeace ⁶ has placed tiger shrimp (*Peneaus monodon*) and Pacific white shrimp (*Litopenaeus vannamei*) in the seafood red list category for unsustainable fishing practices involved. This means the western food fish market is limiting their purchase of tropical prawns, hence adversely affecting our coastal aquaculture commerce. Intensive stocking and excessive feed results in stress

and disease related loss. White spot syndrome is a relevant example in this context^{7,8}.

An additional concern is the use of antibiotics which are causing mutations within the microbial gene pool. FAO strongly recommends against the use of such drugs because of grave future impacts. Use of such chemicals is affecting human health via chemical residues accumulated in fish. The impact has serious consequences for the future. Multiple drug resistant strains of microorganisms are evolving creating imbalance in the ecosystem. Moreover uses of antibiotics also destroy beneficial microbes which boost aquaculture performance⁹.

Feed is of prime importance in modern aquaculture and accounts for a major share of total operational cost. Better utilization of feed directly affects profitability and environmental sustainability¹⁰. Fish meal has historically been the protein source of choice in aquatic feeds, but global supplies have reached a plateau making it less available and more expensive¹¹. Cost reduction of feed is a crucial aspect for the sustainability of the aquaculture sector. This can be achieved by reducing the amount of the most expensive feed constituent with an alternative ingredient without compromising on quality and yield; and poses no threat to the environment. Protein is the most expensive commodity in feed and fish meal is still the protein source held in very high regard. Fish meals have their own set of concerns: 1) disease vectors bringing infection into the culture pond, 2) stagnant global production volumes and 3) inconsistent supply based on seasonal fluctuations. Excess feeding is another issue since farmers administer large quantity to make up for the lower nutritional value of feeds¹². In order to help the fish cultivators and address this concern, formulated feeds with minimal ecological impacts could be the mainstay.

Gradually the interest is moving towards plant based alternative protein sources. Apart from soyabean meal, floral mangrove associates native to Sundarbans such as salt marsh grass (*Porteresia coarctata*), and green algae (*Ulva lactuca*, *Enteromorpha intestinalis*) rich in protein and carotenoids are being used to develop meals for prawn cultivation with success^{13,14}. Such endeavors bring to light alternative and sustainable sources for environment friendly aquaculture. Unfortunately, the use of cheaper crude proteins from mustard cakes, dehydrated trash fish and animal entrails has flourished within the aquaculture feed industry^{15,11} which has resulted in deterioration of both fresh and brackishwater ponds leading to diseases, compromised yield and ecological damage^{16,17}.

Modern sustainable nutrition has been successfully used to improve immunity and help counter diseases, such as use of feed additives like β -glucans from yeast to boost fish immunity reducing disease occurrence in tropical prawns^{18,19,20}. Immunity boosting nutrition is the buzz word, especially with the health impacts of antibiotics. Nucleotide supplementation in pacific white shrimp shows improved resistance against white spot syndrome virus, WSSV²¹. Similarly the use of alginates (from marine algae) stimulates the immune system in tropical and moderate climate fishes²². Use of probiotics is showing promising results too in improved health, growth and feed conversion ratio^{23,24,25}.

However not all immunostimulants can be effectively commercialized. High manufacturing costs make feed developers choose from locally available and easily sourced ingredients. Gujarat is the leading producer of commercial marine algae²⁶ and its easy availability can assist us to formulate a combination of garlic and marine algae pure extracts. Amino acids purified from both soyabean and sea weed may be further used as a supplement.

FAO advises through technical investigations the development and implementation of amino acids, natural antibiotic-alternatives and immunostimulant based feed supplementations¹⁵. From 2012 – 2013 West Bengal produced 1490.01 thousand tonnes of fish, ranking them the second largest fish producing state after Andhra Pradesh²⁷. Adopting eco-friendly formulations in fishing areas of West Bengal may result in improving our fish trade. Although we can discuss at length about the advantages of alternative feed supplements, the primary challenge is to motivate the fish-farmers towards adopting such technologies. The paper provides information on the properties of amino acids, allicin and sea weed extract popular in the aquafeed trade and management action plans for encouraging sustainability in fishing areas of the Indian mangrove Sundarbans where over 70% of the population is dependent on the aquaculture trade. Awareness campaigns in the mangrove belts may assist the coastal communities to adopt more eco friendly strategies to improve fishing and sustain our environment as well as prepare us for an ever increasing demand for food. Extensive peer reviewed articles clearly indicate that amino acids, garlic and marine algae components are effective (Figure 1), and as later discussed there are a few developers pitching for the cause. The challenge is to provide support for the integration of sustainability in aquaculture programmes in ecologically critical areas of the Indian Sundarbans.

AMINO ACIDS

Amino acids can be segmented into essential, non essential and conditionally essential. The essential amino acids must be included in fish diet. The non essential ones can be synthesized by fish when required by using essential amino acids as a substrate. The conditionally essential amino acids are a branch of the non essential category, however they are included in the diet when the rate of their uptake exceeds rate of synthesis²⁸. Amino acids control various biological processes in fish which govern their survival, development, immunity, appetite and health and their deficiency shows a lack of these. They are also documented to work as chemo-attractants of feed, reduce aggressive tendencies and make them tolerant to environmental changes (salinity/temp/pH fluctuations), apart from improving taste of fish. Combinations of both essential and non-essential amino acids have been shown to possess crucial functions in fish physiology. In the current feed development industry, amino acids are seen as either a part of pro-environmental or functional feeds/supplements. The former promotes growth and immunity, while the latter makes the stock tolerant to changes in the environment^{29, 30, 28}. Amino acid supplementation with soya bean meal as a complete replacement of fishmeal is effective in weight gain of juvenile prawns³¹, Nile tilapia³², and rainbow-trout^{33, 34} suggesting a possibility of a reduction in protein based meals for fish without compromising on growth rate and hence reducing operation costs. Alanine and glycine have been demonstrated to improve appetite and act as chemo attractants for juvenile fishes³⁵. Arginine has been seen to promote a variety of functions in different fishes. In channel catfish it inhibits pathogenic bacteria³⁶; in tilapia it controls neural development³⁷. Supplementing the diet of hybrid striped bass with either arginine or glutamine has shown to boost the immune system by increasing superoxide anion content of macrophages. It also improves growth and gut development³⁸. A combination supplementation with arginine and histidine greatly improves growth performance in Olive flounder³⁹. Cysteine, glutamic acid and glycine acts as antioxidants by synthesizing glutathione in all fishes⁴⁰. Sulphur containing amino acids methionine and cysteine are crucial for gene expression and protein synthesis⁴¹. Methionine supplemented with plant based feed exhibit desired growth in Pacific white shrimp⁴². Glutamine has been shown to promote growth and intestine development together with improved food digestion in carp species⁴³. Histidine has been documented to make salmon tolerant to alterations in pH⁴⁴ and proline promotes

growth⁴⁵. Arginine and methionine promote growth in tiger prawns^{46, 47}; while threonine promotes adequate weight gain in Pacific white prawn⁴⁸. Tryptophan supplementation shows suppression of aggressive behavior in juvenile cod⁴⁹ and improved high temperature tolerance with reduced oxygen consumption in Indian carp *Cirrhinus mrigala*⁵⁰. Valine supplementation has been documented to alter the gut flora composition and improve feed absorption and digestion coupled with growth in Jian carp⁵¹. Leucine improves the immune system while lysine and methionine facilitate lipid transport in fishes^{52, 53}. Tyrosine and phenylalanine are known to improve metamorphosis in fish larvae as they improve thyroid hormone synthesis⁵⁴ and have been shown to promote growth in milk fish⁵⁵. Supplementation with lysine and methionine in soyabean fed milk fish and grey mullet demonstrates optimum growth with minimal metabolic wastes⁵⁶. All the various amino acids have certain functions in fish physiology and they meet both functional and environmental needs of fishes. In Figure 2 we consider the various amino acids and their effectiveness in sustaining our aquaculture demands.

ALLICIN

With the growing demand for fish, the aquaculture sector is maximizing their production by using excess feed formulations with high stocking density. Such practices are resulting in immune compromised stock resulting in bacterial and fungal infections. To counter disease related loss, various antibacterial chemicals have been applied which are causing complications. Health of cultured fish is deteriorating together with a threat to the environment and humans. Bacterial antibiotic resistance is a major concern. As a result, safe performance boosting alternatives are a must. Garlic has been celebrated for centuries as a natural immune-stimulant and antimicrobial agent, dating back to ancient Egyptian medicine⁵⁷. Garlic's zero side effects on our ecology and health makes it a choice pick for sustainable feed development.

The active ingredient of garlic is predominantly thiosulfinates called Allicin. Its broad spectrum antibacterial, antifungal and antiviral properties have been extensively documented. This includes drug resistant *E.coli* strains, protozoans and platyhelminthes which affect *Lates calcarifer* farming. This exclusive protection is attributed to its reaction to thiol groups of core biological enzymes like RNA polymerase^{58, 59, 60}. However, researchers reveal the selective action of garlic where it acts against pathogenic variety and improves performance of beneficial ones like *Lactobacillus bifidus*⁶¹.

Researchers²⁴ discuss the probiotic feed benefits of this species of bacteria in aquaculture, suggesting that garlic extracts can be beneficial in enhancing such formulations. In Thailand, garlic is being mixed into feed and administered to prawn farms to avoid bacterial infections⁶².

Together with garlic's excellent track record in controlling pathogens, it's been observed to boost growth, weight gain and blood content in fishes. Overall fish growth and haemoglobin has been documented to be significantly higher than controls⁶³. In tilapia the supplementation of garlic has improved survival rates by stimulating a spike in monocytes. Garlic improved storage life of flesh, as it was demonstrated to last longer in ice with minimal mould infection⁵⁷. Improved quality of flesh with garlic supplementation has been attributed to an increase in protein content and lowering of lipids⁶⁴. These effects are thought to be due to the rise in glutathione peroxidase which in turn boosts antioxidants hence protecting the tissue against oxidative radicals⁶⁵. Sulphur containing compounds in fish impart its flavour. Investigators⁶¹ reveal that the sulphide components of garlic when included in fish feed can improve both quality and flavour of the flesh. The garlic flavour also acts as a potent feed attractant and improves feed conversion ratio.

SEA WEED EXTRACTS

The importance of sea weeds as functional food is primarily because of their rich source of bioactive polysaccharides, antioxidants and amino acids. Apart from their use as alginate in various food grade and laboratory materials, they have a big role to play in sustainable aquafeed technology.

It is beneficial to use pure extracts rather than sea weeds themselves for the following reasons i) improve bioavailability of its active components, and ii) remove toxins present in the raw source.

Sulfated polysaccharides retard growth of pathogenic bacteria. Studies⁶⁶ show disease resistance against strong pathogens like *Aeromonas hydrophila* in *Pangasianodon* sp. Like allicin they also act as prebiotics. The most celebrated of these are alginates, fucoidan, laminarin and galactans. These polysaccharides compose the cell wall and inter cellular matrix of seaweeds. Fucoidan is sourced from brown seaweed and possess anti viral, anti tumour, anti inflammatory and antioxidant properties. Scientists⁶⁷ have demonstrated increased survival rate of white spot syndrome virus affected tiger prawns by administering fucoidan extracts from brown or green seaweed. Laminarin is also present in brown seaweed had acts as a prebiotic, apart from containing anti viral and antibiotic properties. Nearly

47% of brown seaweed biomass is comprised of alginate and acts as a potent antibiotic and anti inflammatory. Galactans have anti tumour and antiviral activities^{68,69}.

Antioxidants present in seaweed are in the form of carotenoids, phycobiliproteins and polyphenols. Both of these are strong free radical scavengers with phycobiliproteins possessing additional antiviral and anti inflammatory values. Pholorotannins are a form of polyphenols which has both antioxidant and antibacterial values^{68,69,22}.

Seaweeds are rich in amino acids aspartic acid and glutamic acid. In red seaweeds they comprise 14 and 19% of total amino acids. While in green seaweeds they are 26 and 32% respectively. The highest content is in brown seaweeds with 22 and 44% of total amino acids⁷⁰. Utilizing mixed sea weed extracts provide feed developers a full profile to work with and improve on the sustainability quotient.

CONCLUSION

The River Ganges flow out into the Bay of Bengal and at this confluence has formed the Indian Sundarban Mangrove ecosystem, which is an UNESCO World Heritage site for its rich biodiversity and productivity. It spans an area of 9630 sq. km and comprises of 102 islands⁷¹. Fishing areas of Kakdwip, near the banks of the Kalnagini River (21°52'06"- 21°86'83"N, 88°11'12"E - 88°18'67"E), Namkhana (21.7667° N, 88.2333° E) and Sagar (21.8° N, 88.1° E) in South 24 Parganas, Indian Sundarbans are been extensively covered (See map in Fig. 3) in an effort to create awareness regarding sustainable aquaculture practices as a large proportion of aquafarmers are engaged in high risk intensive monoculture of tropical shrimps and prawns like *Penaeus monodon* and *Litopenaeus vannamei*. Suggested methods: (a) conducting education seminars through local Block Development Offices (BDOs) to current and prospective brackishwater cultivators as well as Government fisheries officers; (b) door to door interaction with aquafarmers; (c) distribution of free samples of supplements to aquafarmers and educating them about the benefits of using such low cost immune boosters along with sustainable practices of polyculture for tropical prawns (See Fig. 4)

The important factor is building trust, and that is done with advertising trials in experimental ponds and by providing samples to aquafarmers through seminars. A sustainable solution has meaning once visible tests are positive and farmers can 'see' the difference. Conducting seminars through Panchayat Block Development Offices is a way of breaking the ice; however it is far from melting it.

The challenge is 'accepting change'. In spite of positive results local farmers are always apprehensive about trying a new solution. Hence local network with the local village development offices is a crucial step towards bridging the gap. Economic development of local fishing communities is a necessary step for overcoming their mind block on accepting a new solution. Various manufacturers have supplied spurious products with false claims that have also contributed to this hindrance¹¹. The strength of the aquaculture community comes from the sustainability quotient. Excess feed inputs can disrupt the balance of the fish trade; however its control with designed nutrition has a promising future⁷². Antibiotics face a strong criticism from the community, and herbal resources are still held in high regard as an alternative option. Local communities have shown considerable interest in herb based products and appreciate the need for alternative supplements which has sparked researchers to continue their investigations on herb based drugs in the feed industry^{64, 73, 74}.

Direct farmer interactions are a must in building this trust. Door to door promotion attracts a fair response from prawn and fish cultivators of the area. Regular customer contact and feed-backs can generate 'need' based solutions. Such strategies may streamline productivity based on precise formulations as per the requirements and conditions. The manufacturer has to alter formulations for the desired result, hence creating a repertoire of feed formulations²⁸. With a gradual shift towards plant protein sources, formulated amino acid nutritional supplementation will further the efforts in increasing efficiency in feed utilization without compromising with metabolic performances⁷⁵.

It is imperative to improve profits for the fish farmer. Cost based analysis is an ongoing process. Pricing and logistics can prove to be a hindrance in farmer support since affordability and timely administration ensures a good crop, hence strategic locations need to be identified which can reach out to remote centres in the Indian Sundarbans⁷⁶.

A few commercial fish cultivators of the Indian Sundarbans start implementation; now the real question is whether such sustainable options will make a difference in lowering the use of excess feed, antibiotics and other potentially harmful chemicals and pave the green path towards saving our mangroves. Herbal and amino acid based feed supplement solutions are currently available to the

Indian aquafarmers like Grofast, Prolive and Biona from SRIBS Biotekno International (www.sribsbio.in); Lucanthin, Procee (contains amla extracts) and Promass from Anfomed India (<http://www.anfomedindia.com/aquaculture.html>); Lividol-FS-Aqua from Neospark (<http://www.neospark.com/aquaculture.html>). Such trends indicate a keen interest in formulators to maintain India's world ranking in aquaculture produce. However, in spite of the design and manufacturing, there is no real presence of these nutritional options in ecologically sensitive aquaculture regions like the Indian Sundarbans in spite of the very fact that eco-friendly solutions forms a stable backbone of this high demand aqua-feed industry where protein is an expensive commodity⁷⁴.⁷⁶ There is no doubt that pharmaceutical antibiotics can deliver in sprint runs, however the future remains obscured. Hence technology is gradually going back to the knowledge of the ancient Ayurvedas, the Unani and Egyptian medicine of herbal uses^{73, 74}. The World Bank survey reveals that in countries of South Asia, per capita income is on the rise, which is resulting in an increase in food fish consumption⁷⁷. Combined with new age biotechnology we can isolate purified bioactive components which are stronger and greener! Moreover, programmes for active culture of marine algae in the Sundarbans are strongly recommended. It is beneficial to use extracts rather than the plants themselves for the following reasons: i) improves bioavailability of its active components, and ii) removes any toxins present in the raw source⁷³. With versatile functions and applications in therapeutic feed, sustainable supplementation holds considerable potential in the growth of modern day aquaculture together with protecting our coastal communities⁷⁶.

ACKNOWLEDGEMENT

I am grateful to the people of Kakdwip, Namkhana and Sagar for their help and understanding. My heartfelt thanks to Shri Purnendu Shekhar Mondal, Director of both the Nagendra D.Ed College for Girls, Rudranagar, Sagar Island and the Namkhana B.Ed College for his support to the cause. I am extremely grateful to the NatureNerve team, Kolkata for sharing the passion for aquaculture sustainability and thankful to Techno India University, Kolkata for the necessary infrastructure and facility.

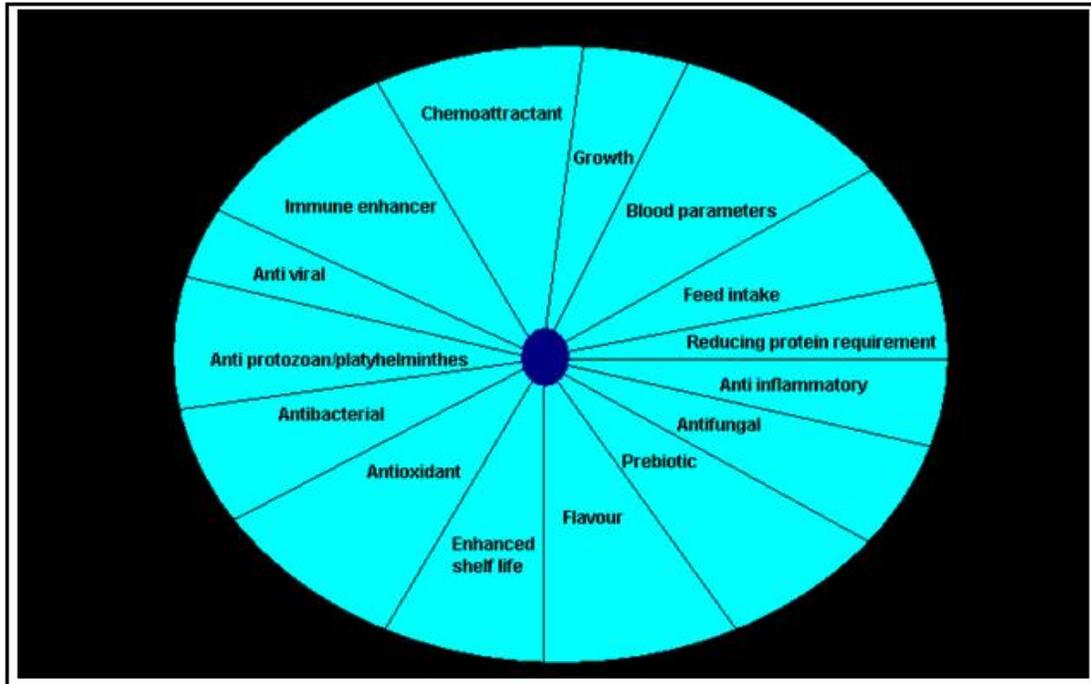


Fig. 1

The properties of amino acids, allicin and sea weed extracts (Li et al, 2009; Lee and Gao, 2012; Jana et al, 2012; Nwabueze, 2012; Militz et al, 2014; Chojnacka et al, 2012).

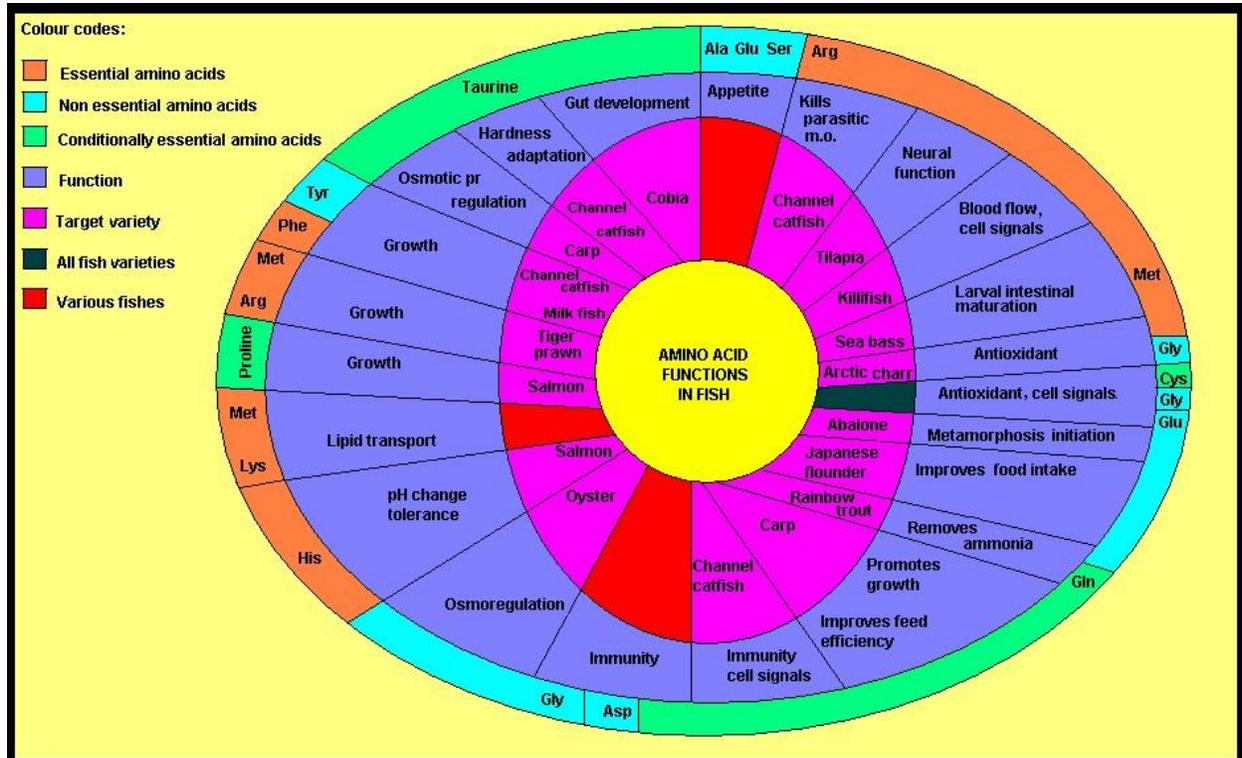


Fig. 2

Properties of amino acid in aquaculture nutrition (adapted from Li et al, 2009).

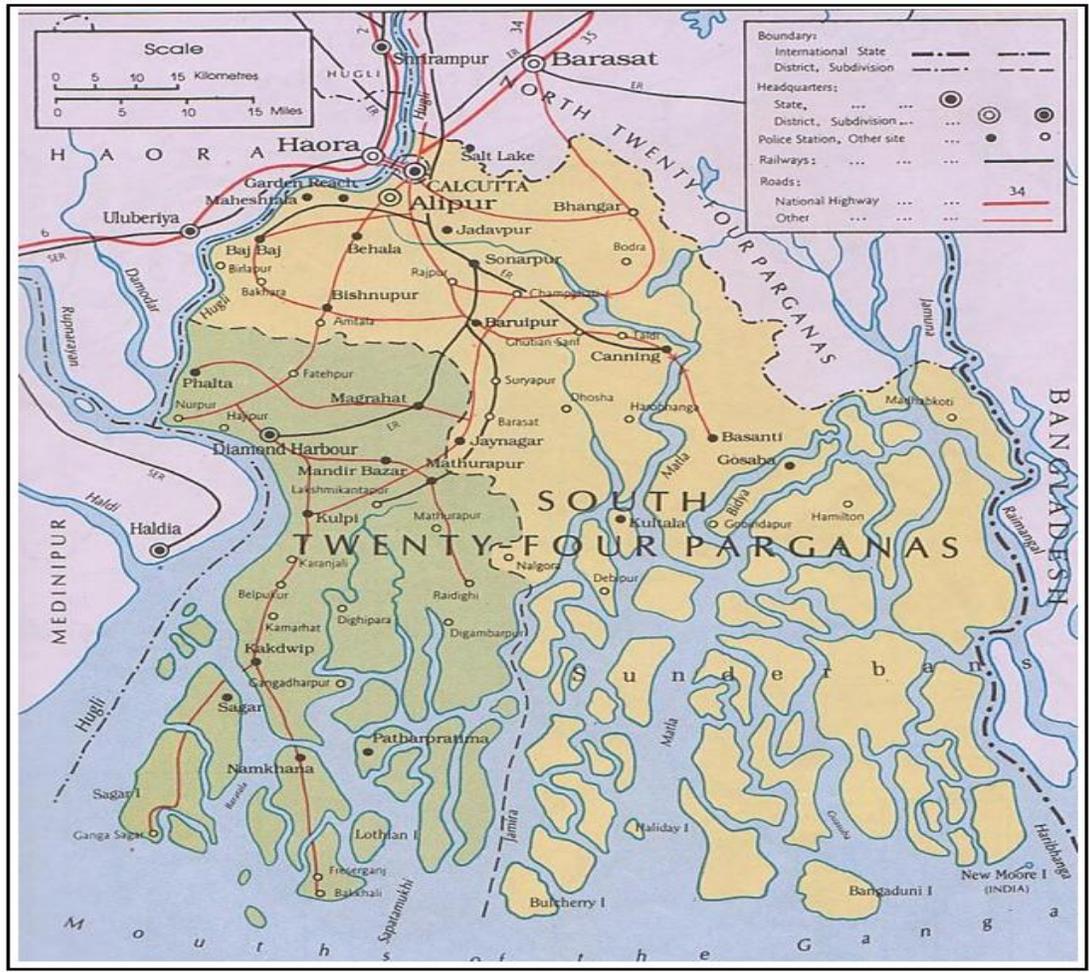


Fig. 3
Map of the Indian Sundarbans with the major towns. Map courtesy: Indian Railways



Fig. 4
120 day culture of *Peneus monodon* using amino acid, allicin and sea weed based feed supplementation in Kakdwip, Indian Sundarbans.

REFERENCES

1. FAO Fisheries and Aquaculture Department. Global Aquaculture Production Statistics for the year 2012. Published 2014; <ftp://ftp.fao.org/FI/STAT/Overviews/AquacultureStatistics2012.pdf>
2. Troell M. Integrated marine and brackishwater aquaculture in tropical regions: research, implementation and prospects. In D. Soto (ed.). Integrated mariculture: a global review. FAO Fisheries and Aquaculture Technical Paper, Rome, FAO, 2009; 529: 47 – 131.
3. Primavera JH. Socio-economic impacts of shrimp culture. *Aquaculture Research*, 1997; 28: 815 – 827.
4. Valiela I, Bowen JL, York JK. Mangrove forests: One of the world's threatened major tropical environments. *BioScience*, 2001; 51(10): 807-815.
5. Thornton C, Shanahan M, Williams J. From wetlands to wastelands: Impacts of shrimp farming. *Wetland Science and Practice*, 2003; 20(1): 48 – 53.
6. Greenpeace International Seafood Red List. <http://www.greenpeace.org/international/en/campaigns/oceans/which-fish-can-I-eat/red-list-of-species/>
7. Karunasagar I, Otta SK, Karunasagar I. Histopathological and bacteriological study of white spot syndrome of *Penaeus monodon* along the west coast of India, *Aquaculture*, 1997; 153: 9 – 13.
8. Lo C, Leu J, Ho C, Chen C, Peng S, Chen Y, Chou C, Yeh P, Huang C, Chou H, Wang C, Kou G. Detection of baculovirus associated with white spot syndrome (WSBV) in penaeid shrimps using polymerase chain reaction, *Diseases of Aquatic Organisms*, 1996; 25: 133 – 141.
9. FAO, OIE and WHO Joint Report. Antimicrobial use in aquaculture and antimicrobial resistance. Seoul, Republic of Korea, 2006; ftp://ftp.fao.org/ag/agn/food/aquaculture_rep_13_16june2006.pdf
10. De Silva SS. Reducing feed costs in aquaculture: Is the use of mixed feeding schedules the answer for semi-intensive practices? *Aquaculture Asia*, 2006; 11(4): 7 – 12.
11. Rana KJ, Siriwardena S, Hasan MR. Impact of rising feed ingredient prices on aquafeeds and aquaculture production. FAO Fisheries and Aquaculture Technical Paper 2009; 541: 63p.
12. Craig S, Helfrich LA. Understanding Fish Nutrition, Feeds, and Feeding. Virginia Tech, 2009; Virginia Cooperative Extension 420-256. https://pubs.ext.vt.edu/420/420-256/420-256_pdf.pdf
13. Mitra A, Banerjee K. Mangroves and their associates: our next door neighbour. WWF-India publishing, 2006; Canning Field Office, Sundarbans Landscape Project
14. Mondal K, Ghosh R, Bhattacharyya SB, Zaman S, Mallik A, Das M, Mitra A. Partial replacement of fish meal with mangrove based plant ingredients and its effect on water quality, growth performance and length-weight relationship of freshwater prawn *Macrobrachium rosenbergii*. *Species*, 2013; 3(8): 15 – 21.
15. Naylor RL, Goldberg RJ, Primavera JH, Kautsky N, Beveridge MCM, Clay J, Folke C, Lubchenco J, Mooney H, Troell M. Effect of aquaculture on world fish supplies. *Nature* 2000; 405: 1017 – 1024.
16. Cho CY, Bureau DP. A review of diet formulation strategies and feeding systems to reduce excretory and feed wastes in aquaculture. *Aquaculture Research*, 2001; 32(1): 349 – 360
17. Webb Jr. KA, Gatlin III DM. Effects of dietary protein level and form on production characteristics and ammonia excretion of red drum *Sciaenops ocellatus*. *Aquaculture*, 2003; 225: 17 – 26.
18. Suphantarika M, Khunrae P, Thanardkit P, Verdyun C. Preparation of spent brewer's yeast -glucans with a potential application as an immunostimulant for black tiger shrimp, *Penaeus monodon*. *Bioresource Technology*, 2003; 88: 55 – 60.
19. Chang CF, Su MS, Chen HY, Liao IC. Dietary -1,3-glucan effectively improves immunity and survival of *Penaeus monodon* challenged with white spot syndrome virus. *Fish & Shellfish Immunology*, 2003; 15: 297 – 310.
20. Ganguly S. Feed supplements for sustainable aquaculture with special emphasis to immunostimulant additives for prevention of disease transmission in aquatic habitats: An editorial. *Research Journal of Chemical and Environmental Sciences*, 2013; 1(3): 1 – 2.
21. Andrino KGS, Serrano Jr AE, Corre Jr VL. Effects of Dietary Nucleotides on the Immune Response and Growth of Juvenile Pacific White Shrimp *Litopenaeus vannamei* (Boone, 1931). *Asian Fisheries Science*, 2012; 25: 180 – 192.

22. Ringo E, Olsen RE, Vecino JLG, Wadsworth S, Song SY. Use of immunostimulants and nucleotides in aquaculture: a review. *Journal of Marine Science Research and Development*, 2012; 2(1): 1 – 22.
23. Rengpipat S, Rukpratanporn S, Piyatiratitivorakul S, Menasaveta P. Immunity enhancement in black tiger shrimp (*Penaeus monodon*) by a probiotic bacterium (*Bacillus S11*). *Aquaculture*, 2000; 191: 271 – 288.
24. Miscevic M, Cirkovic M, Jovanovic R, Ljubojevic D, Novakov N, Masic Z, Markovic M. Effect of probiotics on the production of one-year old tench and common carp. *Archiva Zootechnica*, 2012; 15(4): 69 – 76.
25. Ghosh AK, Sarkar S, Bir J, Islam SS, Huq KA, Naser MN. Probiotic Tiger Shrimp (*Penaeus monodon*) Farming at Different Stocking Densities and its Impact on Production and Economics. *International Journal of Research in Fisheries and Aquaculture*, 2013; 3(2): 25 – 29.
26. Mitra A, Banerjee K, Basu S, Mukherjee A, Mandal B. Training manual on non-classical uses of mangrove resources of Indian Sundarbans for alternative livelihood programmes: Use of seaweeds in fishfeed preparation. WWF-India publishing, 2006; Canning Field Office, Sundarbans Landscape Project.
27. Ministry of Agriculture, Government of India. Fisheries profile of India. Dept of Animal Husbandry, Dairying & Fisheries, 2013; <http://www.dahd.nic.in/dahd/WriteReadData/Fisheries%20Profile%20of%20India%2015.7.2013.pdf>
28. Li P, Mai K, Trushenski J, Wu G. New developments in fish amino acid nutrition: towards functional and environmentally oriented aquafeeds. *Amino Acids*, 2009; 37(1): 43 – 53.
29. Ketola GH. Amino acid nutrition of fishes: Requirements and supplementation of diets. *Comparative Biochemistry and Physiology*, 1982; 73B(1): 17 – 24.
30. Cowey CB. Amino acid requirements of fish: a critical appraisal of present values. *Aquaculture*, 1994; 124: 1 – 11.
31. Floreto EAT, Bayer RC, Brown PB. The effects of soybean-based diets, with and without amino acid supplementation, on growth and biochemical composition of juvenile American lobster, *Homarus americanus*. *Aquaculture*, 2000; 189: 211 – 235.
32. Furuya WM, Pezzato LE, Barros MM, Pezzato AC, Furuya VRB, Miranda EC. Use of ideal protein concept for precision formulation of amino acid levels in fish-meal-free diets for juvenile Nile tilapia (*Oreochromis niloticus* L.). *Aquaculture Research*, 2004; 35: 1110 – 1116.
33. Rodehutsord M, Becker A, Pack M, Pfeffer E. Response of rainbow trout (*Oncorhynchus mykiss*) to supplements of individual essential amino acids in a semipurified diet, including an estimate of the maintenance requirement for essential amino acids. *The Journal of Nutrition*, 1997; 127(6): 1166 – 1175.
34. Gaylord TG, Barrows FT. Multiple amino acid supplementations to reduce dietary protein in plant-based rainbow trout, *Oncorhynchus mykiss*, feeds. *Aquaculture*, 2009; 287: 180 – 184.
35. Shamushaki VAJ, Kasumyan AO, Abedian A, Abtahi B. Behavioural responses of the Persian sturgeon (*Acipenser persicus*) juveniles to free amino acid solutions. *Marine and Freshwater Behaviour and Physiology*, 2007; 40: 219 – 224.
36. Pohlenz C, Buentello A, Helland SJ, Gatlin III DM. Effects of dietary arginine supplementation on growth, protein optimization and innate immune response of channel catfish *Ictalurus punctatus* (Rafinesque 1818). *Aquaculture Research*, 2014; 45(3): 491 – 500. doi:10.1111/j.1365-2109.2012.03252.x
37. Bordieri L, di Patti MCB, Miele R, Cioni C. Partial cloning of neuronal nitric oxide synthase (nNOS) cDNA and regional distribution of nNOS mRNA in the central nervous system of the Nile tilapia *Oreochromis niloticus*. *Molecular Brain Research*, 2014; 142: 123 – 133.
38. Cheng Z, Gatlin III DM, Buentello A. Dietary supplementation of arginine and/or glutamine influences growth performance, immune responses and intestinal morphology of hybrid striped bass (*Morone chrysops* × *Morone saxatilis*). *Aquaculture*, 2012; 362 – 363: 39 – 43.
39. Han Y, Koshio S, Ishikawa M, Yokoyama S. Interactive effects of dietary arginine and histidine on the performances of Japanese flounder *Paralichthys olivaceus* juveniles. *Aquaculture*, 2013; 414 – 415: 173 – 182.
40. Wu G, Fang YZ, Yang S, Lupton JR, Turner ND. Glutathione metabolism and its implications for health. *Journal of Nutrition*, 2004; 134:489–492.
41. Tesseraud S, Coustard SM, Collin A, Seiliez I. Role of sulfur amino acids in controlling nutrient metabolism and cell functions:

- implications for nutrition. *British Journal of Nutrition*, 2009; 101: 1132 – 1139.
42. Fox JM, Humes M. Evaluation of Methionine Supplements and Their Use in Grain-based Feeds for *Litopenaeus vannamei*. *Journal of the World Aquaculture Society*, 2011; 42(5): 676 – 686.
 43. Lin Y, Zhou X. Dietary glutamine supplementation improves structure and function of intestine of juvenile Jian carp (*Cyprinus carpio* var. Jian). *Aquaculture*, 2006; 256: 389 – 394.
 44. Mommsen TP, French CJ, Hochachka PW. Sites and patterns of protein and amino acid utilization during the spawning migration of salmon. *Canadian Journal of Zoology*, 1980; 58: 1785 – 1799.
 45. Aksnes A, Mundheim H, Toppe J, Albrektsen S. The effect of dietary hydroxyproline supplementation on salmon (*Salmosalar* L.) fed high plant protein diets. *Aquaculture*, 2008; 275: 242 – 249.
 46. Liao IC, Liu FG. A brief review of nutritional studies for *Penaeus monodon*. *Advances in Tropical Aquaculture*, 1989; 9: 355 – 380.
 47. Chen HY, Leu YT, Roelants I. Quantification of arginine requirements of juvenile marine shrimp, *Penaeus monodon*, using microencapsulated arginine. *Marine Biology*, 1992; 114: 229 – 233.
 48. Zhou QC, Wang YL, Wang HL, Tan BP. Dietary threonine requirements of juvenile Pacific white shrimp, *Litopenaeus vannamei*. *Aquaculture*, 2013; 392 – 395: 142 – 147.
 49. Hoglund E, Bakke JM, Overli O, Winberg S, Nilsson GE. Suppression of aggressive behaviour in juvenile Atlantic cod (*Gadus morhua*) by L-tryptophan supplementation. *Aquaculture*, 2005; 249: 525 – 531.
 50. Tejpal CS, Sumitha EB, Pal AK, Murthy HS, Sahu NP, Siddaiah GM. Effect of dietary supplementation of L-tryptophan on thermal tolerance and oxygen consumption rate in *Cirrhinus mrigala* fingerlings under varied stocking density. *Journal of Thermal Biology*, 2014; 41: 59 – 64.
 51. Dong M, Feng L, Kuang SY, Liu Y, Jiang J, Hu K, Jiang WD, Li SH, Tang L, Zhou XQ. Growth, body composition, intestinal enzyme activities and microflora of juvenile Jian carp (*Cyprinus carpio* var. Jian) fed graded levels of dietary valine. *Aquaculture Nutrition*, 2013; 19: 1 – 14.
 52. Li P, Gatlin DM. Evaluation of dietary supplementation of -hydroxy- -methylbutyrate for hybrid striped bass *Morone chrysops* X *Morone saxatilis*. *Journal of Applied Aquaculture*, 2007; 19: 77 – 88.
 53. Harpaz S. L-Carnitine and its attributed functions in fish culture and nutrition—a review. *Aquaculture*, 2005; 249: 3 – 21.
 54. Pinto W, Figueira L, Dinis MT, Aragao C. How does fish metamorphosis affect aromatic amino acid metabolism? *Amino Acids*, 2009; 36: 177 – 183.
 55. Borlognan IG, Coloso RM. Requirements of juvenile milkfish (*Chanos chanos* Forsskal) for essential amino acids. *Journal of Nutrition*, 1993; 123(1): 125 – 132.
 56. Jana SN, Sudesh, Garg SK, Garg SK, Sabhlok VP, Bhatnagar A. Nutritive evaluation of Lysine- and Methionine supplemented raw Vs heat-processed soybean to replace fishmeal as a dietary protein source for grey mullet, *Mugil cephalus*, and milkfish, *Chanos chanos*. *Journal of Applied Aquaculture*, 2012; 24: 69 – 80.
 57. Aly SM, Atti NMA, Mohamed MF. Effect of garlic on the survival, growth, resistance and quality of *Oreochromis niloticus*. *International Symposium on Tilapia in Aquaculture*, 2008; 8: 277 – 296.
 58. Ankri S, Mirelman D. Antimicrobial properties of allicin from garlic. *Microbes and Infection*, 1999; 1: 125 – 129.
 59. Militz TA, Southgate PC, Carton, AG, Hutson KS. Dietary supplementation of garlic (*Allium sativum*) to prevent monogenean infection in aquaculture. *Aquaculture*, 2013; 408 – 409: 95 – 99.
 60. Militz TA, Southgate PC, Carton AG, Hutson KS. Efficacy of garlic (*Allium sativum*) extract applied as a therapeutic immersion treatment for *Neobenedenia* sp. management in aquaculture. *Journal of Fish Diseases*, 2014; 37(5): 451 – 461.
 61. Lee JY, Gao Y. Review of the application of garlic, *Allium sativum*, in aquaculture. *Journal of the World Aquaculture Society*, 2012; 43(4): 447 – 458.
 62. Direkbusarakom S. Application of medicinal herbs to aquaculture in Asia. *Walailak Journal of Science & Technology*, 2004; 1(1): 7 – 14
 63. Nwabueze AA. The effect of garlic (*Allium sativum*) on growth and haematological parameters of *Clarias gariepinus* (Burchell, 1822). *Sustainable Agriculture Research*, 2012; 1(2): 222 – 228.
 64. Xiang X, Liu CZ. Effect of allicin on growth of *Colossoma barchipomum*. *Fisheries Science*

- and Technology Information, 2002; 29: 222 – 225.
65. Metwally MAA. Effects of garlic (*Allium sativum*) on some antioxidant activities in tilapia Nilotica (*Oreochromis niloticus*). World Journal of Fish and Marine Sciences 2009; 1(1): 56 – 64.
 66. Prabu DL, Sahu NP, Pal AK, Dasgupta S, Narendra A. Immunomodulation and interferon gamma gene expression in sutchi cat fish, *Pangasianodon hypophthalmus*: effect of dietary fucoidan rich seaweed extract (FRSE) on pre and post challenge period. Aquaculture Research, 2014; 1 – 20.
 67. Chotigeat W, Tongsupa S, Supamataya K, Phongdara A. Effect of fucoidan on disease resistance of black tiger shrimp. Aquaculture, 2004; 233: 23 – 30.
 68. Holdt SL, Kraan S. Bioactive compounds in seaweed: functional food applications and legislation. Journal of Applied Phycology, 2011; 23: 543 – 597.
 69. Chojnacka K, Saeid A, Witkowska Z, Tuhy L. Biologically active compounds in seaweed extracts - The prospects for the application. The Open Conference Proceedings Journal, 2012; 3(1M4): 20 – 28.
 70. Fleurence J. Seaweed proteins: biochemical, nutritional aspects and potential uses. Trends in Food Science & Technology, 1999; 10: 25 – 28
 71. Chaudhuri AB, Choudhury A. Mangroves of the Sundarbans India. Published by IUCN, 1994.
 72. Troell M, Naylor RL, Metian M, Beveridge M, Tyedmers PH, Folke C, Arrow KJ, Barrett S, Crepin A-S, Ehrlich PR, Gren A, Kautsky N, Levin SA, Nyborg K, Osterblom H, Polasky S, Scheffer M, Walker BH, Xepapadeas T, Zeeuw A. Does aquaculture add resilience to the global food system? Proceedings of the National Academy of Sciences, 2014; 111(37): 13257 – 13263.
 73. Asimi OA, Sahu NP. Herbs/spices as feed additive in aquaculture. Scientific Journal of Pure and Applied Sciences, 2013; 2(8): 284 – 292.
 74. Ramudu KR, Dash G. A review on herbal drugs against harmful pathogens in aquaculture. American Journal of Drug Discovery and Development, 2013; 3(4): 209 – 219.
 75. Nunes AJP, Sa MVC, Browdy CL, Vazquez-Anon M. Practical supplementation of shrimp and fish feeds with crystalline amino acids. Aquaculture, 2014; 431: 20 – 27.
 76. Chinarong A, Krishna BY. Trends and progress of the aqua feed companies in India a case of Pancharathna Companies. International Journal of Scientific and Research Publications, 2013; 3(6): 1 – 7.
 77. The World Bank. Fish to 2030: Prospects for fisheries and aquaculture. Agriculture and Environmental Services Discussion Paper, 2013; 3(83177-GLB): 1 – 102. www.fao.org/docrep/019/i3640e/i3640e.pdf