

**INTERNATIONAL JOURNAL OF ADVANCES IN  
PHARMACY, BIOLOGY AND CHEMISTRY**

**Research Article**

**Quantitative and Immunohistochemical analysis of  
lymphoid cells of swine Immunized with *Escherichia  
coli* bacteria**

**Enver Zenku<sup>1</sup>, Sheval Memishi<sup>1\*</sup> and Ljiljana Simonovska<sup>2</sup>.**

<sup>1</sup> State University of Tetova, Faculty of Natural Science, Department of Biology,  
Rr. Ilindenit, 1200 Tetovo, Macedonia.

<sup>2</sup> University "Goce Delcev" Stip, Macedonia, Faculty of Medicinal Sciences, Macedonia.

**ABSTRACT**

In this study we have demonstrated lymphoid cells in the small intestine, spleen and mesenteric lymph node (MLN) of the 4 weeks old pigs, crossbreeds between swedish Landras, Hypor and Seghers, after immunization with nonenterotoxigenic (non – ETEC) *Escherichia coli* strain.

Using the immunohistochemical ABC method, with the monoclonal antibodies MIL 13, specific for the CD45RA marker and MIL5, specific for the CD45RC marker, we showed the histological distribution of the CD45RA, and CD45RC lymphoid cells in MLN and small intestine. In the MLN of immunized pigs CD45RA cells are distributed in large numbers in both cortex and paracortex, in the small intestine we found accumulations in the lamina propria and individual cells in the epithel. CD45RC cells are few in the small intestine, in the paracortex of the MLN they are numerous, and only few in the cortex and the sinuses.

Quantitative data for MLN, spleen and the Peyer's patches of ileum were obtained from the flow cytometer. CD45RA marker was found in the larger percentages in the experimental group than in the control group in all of the above organs. Also, CD45RC marker was found in larger percentages in MLN and spleen, while in the Peyer's patches of the experimental group it was in smaller percentage when compared to the control group.

These results indicate increase in the percentage of the lymphoid cells in all of the mentioned organs of immunized animals, and therefore possible immunogenicity of the non – ETEC *E. coli* strain.

**Keywords:** Lymphoid cells, Swine, Immunization and *Escherichia coli*.

**1. INTRODUCTION**

Cells that are participating in the immune system, leukocytes, mobile units present in the body's defense system<sup>5, 7</sup>. Some of them are formed in the bone marrow (granulocytes, monocytes and small number of lymphocytes), a part of lymphatic tissue (lymphocytes and plasma cells). In normal blood there are 6 different types of leukocytes<sup>8, 13, 15</sup>, which are: polymorphonuclear neutrophils, eosinophils, polymorphonuclear, polymorphonuclear basophils, monocytes, lymphocytes, plasma cells and more platelets that are part of the 7th kind of leukocytes which are found in marrow bone-megakariocyte. All of its three kinds of polymorphonuclear leukocytes

have a granular structure for what are called granulocyte. Granulocytes and monocytes protect the body from disease causes phagocytosis, ie with their ingestion. T and B Establish gland lymph organs, thymus and bone marrow. From here lymphocytes pass in peripheral lymphoid organs: spleen, lymph nodes and Peyer's patches the ileum where it ends their differentiation. Developed two similar immune systems, cellular immunity which react with microorganisms immunocompetent cells, foreign cells (tumor or transplantation). For this immunity responsible are T lymphocytes. The other type is the humoral immunity, which focuses on blood

glycoproteins, which are called antibodies that inactivate or eliminate foreign agents<sup>3,4</sup>. Antibodies (immunoglobulins) are blood plasma glycoprotein that relate to antigenic determinants which encourage their creation and those I share in some classes (IgG, IgA, IgM, IgE, IgD)<sup>1,2,10,16</sup>. Some cells suckle IL similar to hormones, which control the activity of cells that are part of the immune reaction<sup>11,14</sup>.

## 2. MATERIALS AND METHODS

In evidence are dealt 7 healthy pigs Swedish Hyporit landrace crucifixion and Segherses age of 4 weeks. The pigs were divided into two groups: 4 in the experimental group, who were immunized with 10 cfu / ac ml. F4 of type Abbotstown in 60 ml triptikaz Bujon strain (TSB) I had with 1.2% NaHCO<sub>3</sub>, 3 animals were equally treat with 60 ml TBS add 1.2% NaHCO<sub>3</sub>. Animals killed 5, 6 or 7-en days after immunization. Was injected intravenously 0.3 ml / kg T61. Immunohistochemical methods and painting ABC - imunohistochemical methods using detected with specific antibodies components that carry antigenic determinants. For the presentation of the reaction between the antigen and antibodies used citochemical reagents. One of the methods is very precise in Histochemistry use of avidin-biotin complex, which is used in all research areas of molecular biology, cell, histology, immunology, clinical biology and pathology. Marking with primare- antibodies of mice used monoclonal antibodies.

## 3. RESULTS

With this research of imunohistochemical through ABC method, is demonstrated localization and distribution of CD 45 RA, CD 45 RC lymph cells in lymph tissue in the digestive system pigs drowned 6 and 7 days after immunization. In the ileum of pigs tested with monoclonal antibodies which marks CD 45 RC, leukocyte cell population appears very rarely CD 45 RC + cells in the intestinal epithelium. In jejunum rarely I find CD 45 RC cells. In the paracortex NLM find large numbers of cells CD 545 RC, and in the region of cortex are rare and lonely, but are also present in the sinuses of the MLC. During acquired quantitative research shows that participation of citometric flow cell CD 45 RA is greater in the ilium Peyer's platches in the group of

experimental animals compared with Peyer plates of the group of control animals, the presence of cells CD 45 RC is smaller Peyer's platches animals in the experiment compared to the control, (Fig.1).

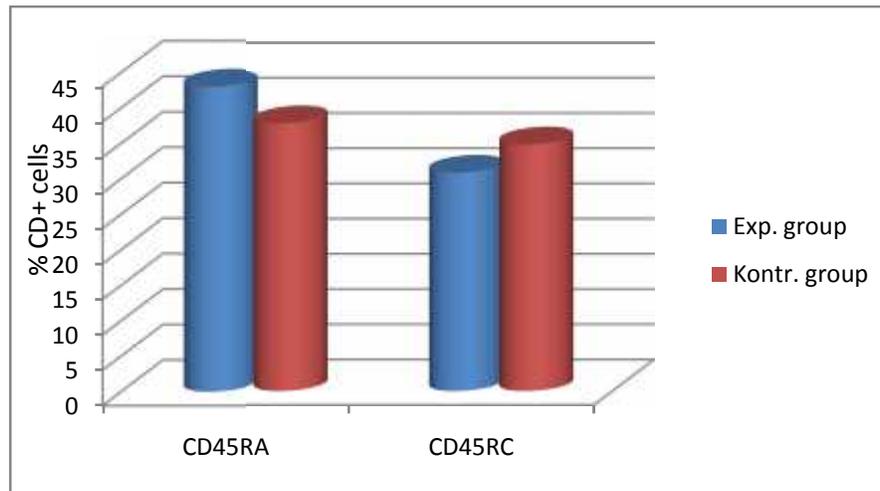
## 4. DISCUSSION

The surface of the mucous membranes of digestive system is under the influence of a large number of different antigens, including harmful substances without food, but also large potential pathogens<sup>9,16,17</sup>. However these differences mechanism is not yet completely clarified, control Conduct for antigens and protection system is achieved by a combination of specific and non-specific protective organisms. Research so far showed that the epithelium of the intestine of swine represents only immune microenvironment<sup>12</sup>. Considerable number of cells present in the epithelium, Peyer's platches, jejunum and ileum are participating in the surveillance of the immune system<sup>6</sup>. Results show increasing presence of CD 45 RC and speeding subpopulation differentiation of T helper.

## 5. CONCLUSION

Based on what was said above, we can conclude that:

1. The tissue distribution of T lymphocytes and lymphoid cells, mijeloide CD 45 and CD 45 RC RA, in mesenteric lymph node (NLM) pigs immunized animal shows of absorption quality issues raised, while the number in lamini proprijji the jejunumit and peyer plates, confirms underdevelopment of the immune system in pigs aged 5 weeks.
2. ABC method proved successful for the existence of lymph cell populations. With optical microscope stained cells differ significantly from those of unstained.
3. Flow cytometry gave us NLM notes, spleen and the peyer plates of the the ilium: CD 45 RA is involved in the largest proportion of animals in the experiment group than the control group of animals. Also marker CD 45 RC is present in greater proportion in the NLM and spleen in the experiment group, while in the ilium PP group in the experiment is expressed as a lower percentage compared with the control group.



**Fig.1.**  
Percentage of lymphoid cells NQ Peyer plates the ilium of the experimental and control animals aged four week.

## 6. REFERENCES

1. Bandrick M, Pieters M, Pijoan C, Baidoo SK, and Molitor TW. Papers: Effect of cross-fostering on transfer of maternal immunity to *Mycoplasma hyopneumoniae* to piglets, *Veterinary Record*, 2011;168(4)100.
2. Bergmann KR, Liu SXL, Tian RL et al., Bifidobacteria stabilize claudins at tight junctions and prevent intestinal barrier dysfunction in mouse necrotizing enterocolitis, *The American Journal of Pathology*, 2013;182(5) 1595–1606.
3. Bozic F, Banovic F, Surlan J, Prevendar Crnic A. Adjuvant activity of levamisole for experimental F18 ac<sup>+</sup> *E. coli* oral vaccine against porcine post-weaning colibacillosis. *Vet. Arhiv*, 2011. 199-209.
4. Bozic F, Cver L, Valpotic I., CD45RA and CD45RC isoforms expression in weaned pigs vaccinated with non-enterotoxigenic F4ac+*Escherichia coli* strain against colibacillosis. *Vet. Med. – Czech*, 2002; 47 (1): 5–11.
5. Clare L and Abram CA. Lowel The Expanding Role for ITAM-Based Signaling Pathways in Immune cells. *Science Signaling*. 2007; 377: p. re2.
6. Del Vecchio G, Tscheik C, Tenz K et al., Sodium caprate transiently opens claudin-5-containing barriers at tight junctions of epithelial and endothelial cells, *Molecular Pharmaceutics*, 2012; 9(9): 2523–2533.
7. Hiroaki H, Tsuneyasu K, Osamu T, Shintaro S, Hideki S, Katsuaki H, Takao H, Hideyuki T, Kiyoshi T, Shizuo A. Small anti-viral compounds activate immune cells *via* the TLR7 MyD88–dependent signaling pathway. *Nature Immunology*. 2002; 3(2): 196 – 200.
8. Horton RE and Vidarsson G. Antibodies and their receptors: different potential roles in mucosal defense, *Frontiers in Immunology*, 2013; 16(4): 200.
9. John MF, Eric N, Carlton LG. *Escherichia coli* in postweaning diarrhea in pigs. *Animal health Research*. 2005; 6(1):17-39.
10. Marcin K, Piotr S, Andreas H, Lin W, Claudia K, Maciej K, Heehyoung L, Anna S, Yong L, Chunmei Y, Jiehui D, Harris S. S., Andrew R., Stephen F., John J R., Drew M. P., Richard J., Hua Y. *In vivo* delivery of siRNA to immune cells by conjugation to a TLR9 agonist enhances antitumor immune responses. *Nature Biotechnology*. 2009; 27(10): 925 – 932.
11. McDole JR, Wheeler LW, McDonald KG et al., Goblet cells deliver luminal antigen to CD103+ dendritic cells in the small intestine, *Nature*, 2012; 483(7389) 345–349.

12. Patel RM, Myers LS, Kurundkar AR, Maheshwari A, Nusrat A, and Lin PW, Probiotic bacteria induce maturation of intestinal claudin 3 expression and barrier function, *The American Journal of Pathology*, 2012; 180(2):626–635.
13. Straw U, D'allaire S, Mengeling, WL, Taylor DI. *Disease of swine*, 8<sup>th</sup> edn. Iowa State University Press, Ames Iowa, USA,. 1999. 799-820.
14. Stuart WN, Christopher L, Thomas EB, Arvin M. Lymfoid follicle dense mucosa at the terminal restum is the principal site of colonization of enterohemorrhagic *Escherichia coli* O157:H7 in the bovine host. *Infect. Immun.* 2003; 71(3); 1505-1512.
15. Yang H., Sallmueller A., Lunnlei JK., Definition of the specificity of monoclonal antibodies against porcine CD45 and CD45R., report from CD45/CD45R and CD44 subgroup of the second international swine CD workshop. *Vet. Immunol. immunopatol.* 2008. 60: 367-387.
16. Zenku E, Memishi SH, Saiti I. Monitoring of Immune Parameters in Patients With Bronchial Asthma in Tetovo Area. *IJSR.* 2014; 3(12):1474-1476.
17. Zenku E, Memishi SH, Haziri A, Identifying of Allergic Agents to Different Human Age Groups in Tetovo Area. *IJSR.* 2014; 3 (8) 2140-2141.