

**INTERNATIONAL JOURNAL OF ADVANCES IN  
PHARMACY, BIOLOGY AND CHEMISTRY**

**Research Article**

**Isolation and characterization of cold-active protease  
producer from ice factory samples**

**Anupama Pathak\*, Narayan Lodge, Joice Gavali and Mukundraj G Rathod.**

School of Life Sciences (DST, FIST & UGC-SAP sponsored),

Swami Ramanand Teerth Marathwada University, Nanded, Vishnupuri, Nanded,

Maharashtra, India - 431 606.

**Abstract**

Two psychrophilic organisms were isolated from the soil sample of ice factory, Phule market, Nanded, Maharashtra. Of these one efficient Cold-active protease producer, casein hydrolysing Gram positive, round shaped motile bacterium was selected and designated as IF1. The isolate was identified by morphological and biochemical characters as *Peptostreptococcus magnus* on the basis of Bergey's manual of systematic bacteriology. It has optimum growth at pH 7 and 4°C temperature. Further extraction, partial purification and characterization of alkaline protease were carried out. After dialysis, catalytic activity of IF1 cold-active protease was determined as 279 IU/ml.

**Keywords:** Ice industry, *Peptostreptococcus magnus*, Protease, Psychrophiles.

**INTRODUCTION**

Proteases refer to a group of enzymes whose function is to catalyse hydrolysis of proteins. Proteolytic enzymes play an important role in digestion as they breakdown the peptide bonds in the protein foods to liberate the amino acids to fulfil the need of body. Beside this, protease has also been used in various forms of therapy and clinical studies like oncology, inflammatory conditions, blood rheology control, and immune regulation<sup>1</sup>.

Detergent fortified with cold active protease can be used in cold wash treatment to improve washing quality of detergent. In present investigation we therefore worked on isolation, characterization of efficient protease producer using ice factory sample<sup>2, 3</sup>.

**MATERIALS AND METHODS**

**A. Sample collection**

Soil samples were collected from an ice factory of Nanded district, Maharashtra. The samples were collected in sterilized polythene bags and transported to the laboratory<sup>4-7</sup>.

**B. Isolation of psychrophiles**

1 gm of composite soil sample was transferred to a 250 mL flask containing 100 mL sterile distilled water and shaken in cooling shaking incubator for 5 °C at 120 rpm for 20 min. Serial dilutions were made and 0.1 mL of aliquots were spread on nutrients agar plates. The plates were incubated for 5 days at 5 °C<sup>8-14</sup>.

**C. Screening of isolate for cold-active proteolytic activity**

Selected isolates were spot inoculated on skimmed milk agar plates and incubated at 5 °C temperature for 5 days. After incubation, plates were observed for the formation of zone of clearance around the colonies<sup>10-14</sup>.

**D. Identification of efficient cold-active protease producer**

Gram staining of selected isolate was performed. Catalase activity, indole production, citrate

utilization, lipase, Pectinase, amylase and Urease production tests were performed. Methyl red, Vogues-Proskauer and carbohydrate fermentation tests were performed by standard procedures. Appropriate positive and negative controls were used<sup>14-20</sup>.

#### E. Cold-active protease production

0.5 mL culture of selected isolate was inoculated in 250 mL of tryptone yeast extract glucose broth containing tryptone 5 g/L, yeast extract 2.5 g/L and glucose 1 g/L (pH 7.2). Protease production was carried out at 5 °C temperature and 100 rpm agitation speed for 48 h in shaking incubator<sup>14-23</sup>.

#### F. Partial purification

The cold active protease was partially purified from fermented broth by centrifuging at 8,944 ×g and 5 °C for 20 minutes. The supernatant was collected and filtered through Whatman filter paper 41µm. To the supernatants ammonium sulphate was added slowly with continuous stirring to the final concentration of 80 % saturation and this solution was allowed to stand for 24 h at 4 °C and further centrifuged at 8,944 ×g for 20 min. The precipitate was resuspended in 50 mM Tris HCl buffer having pH 8.0 and further precipitated with acetone by adding slowly to the final concentration 80 % saturation and left for 1 h at 4°C. The pellet was collected by centrifugation at 8,944 ×g for 20 min at 5°C and resuspended in 20 mM Tris HCl buffer (pH 8.0) and dialyzed against 500 ml of same buffer containing 1 mM MgCl<sub>2</sub> over night at 5 °C with stirring conditions. The dialysate was centrifuged at 5,724 ×g for 20 min at 5 °C and supernatant was collected<sup>14-23</sup>.

#### G. Determination of protein content and proteolytic activity assay

The proteolytic activity was determined by using casein as substrate. The substrate contained 3.75 ml of 1.0 % casein in 100 mM Tris-HCl buffer and 1 mM MgCl<sub>2</sub> at pH 8.0. The 0.5 ml of protease sample was added to the substrate and incubated for half an hour at 5 °C. After incubation, the reaction was quenched with 0.5 mL of 10 % trichloro acetic acid. The quenched reaction mixture was centrifuged at 10,000 rpm for 10 min to pellet precipitated protein and absorbance for the supernatant was determined at 280 nm. One unit of proteolytic was defined as the amount of enzyme that produced an increase of absorbance at 280 nm of 0.1 under the conditions of the assay<sup>14-24</sup>.

## RESULTS AND DISCUSSION

### Isolation, screening and identification

Twelve colonies were appeared on nutrient agar plates. Of these, two morphologically distinct colonies were selected and designated as IF1 and IF2. The extracellular protease producer was screened by streaking bacterial strains on skimmed milk agar and incubated for 48 hrs 4°C.

The isolate IF1 has showed largest size of zone of clearance than IF2. Therefore IF1 isolate was selected for further studies.

IF1 isolate was Gram positive, motile and round shaped bacterium and positive for Catalase, Methyl red, citrate utilization and casein hydrolysis and negative for Indole production, VP, Urease, Cellulase and Pectinase production (Table 2).

It does not utilize any of the fructose, glucose, maltose and sucrose as a carbon source.

Morphological and biochemical characters of IF1 isolate have been given in Table1 and Table 2 respectively.

On the basis of biochemical characterization the isolates IF1 was identified as *Peptostreptococcus magnus* by comparing with standard strain of Bergeys Manual of systematic bacteriology.

### Protease production and assay

The *Peptostreptococcus magnus* secretes large extracellular enzyme protease in the surrounding medium.

The 80% ammonium sulphate saturation leads the precipitation of the protease at 4°C and fractional precipitation with acetone. The excess salt removed from protease by means of a dialysis. The proteolytic activity was determined by using casein as substrate in Tris HCl buffer. Catalytic activity of IF1 protease was determined 279 IU/ml.

## CONCLUSION

Efficient protease producer was isolated from soil sample of Ice factory and identified as *Peptostreptococcus magnus* based on morphological, microscopic, biochemical and physiological characters. *Peptostreptococcus magnus* can be used for production of protease and further in enzyme biotechnology field.

## ACKNOWLEDGMENT

Authors are thankful to Hon'ble Vice Chancellor, S.R.T.M. University, Nanded for providing infrastructure and necessary facilities.

**Table 1**  
**Colony Characters of IF1**

Characters	Isolate IF1
Size	1 mm
Shape	Circular
Color	Off white
Margin	Entire
Surface	Smooth
Elevation	Raised
Consistency	Sticky
Opacity	Translucent

**Table 2**  
**Biochemical Characteristics of IF1**

Test	Result	Test	Result	Test	Result
Catalase	+	Amylase	+	Protease	+
Indole production	-	Urease	-	Fructose	-
Methyl red	+	Lipase	+	Glucose	-
VP	-	Cellulase	-	Maltose	-
Citrate utilization	+	Pectinase	-	Sucrose	-

## REFERENCES

- Rao MB, Aparna M, Tanksale M, Ghatge Sand, Deshpande VV. Molecular and biotechnological aspects of microbial proteases. *Microbiol Mol Biol Rev*, 1998; 62(3): 597–635.
- Dambmann C and Aunstrup K. The variety of serine proteases and their industrial significance In: Turk V, Vitale L, eds. *Proteinases and their inhibitors. Structure, function and applied aspects*. Oxford: Pergamon 1981; 231-244.
- Joshi S, Satyanarayana T. *Biotechnology of Cold-Active Proteases* Biology, 2013; 2(2): 755-783.
- Pathak AP, Rathod MP. Exploration of Unkeshwar hot springs in Maharashtra for thermostable amylase producer. *Res. Rev. Biosci.* 2014; 8(7):269–276.
- Pathak AP, Rathod MG and Rampurkar VV. An ecofriendly approach for thermo stable amylase production using *Bacillus firmus* APP6: a hot spring isolate. *Asiatic J. Biotech. Res.* 2014; 4(4): 101-10.
- Pathak AP and Deshmukh KB. Alkaline protease production extraction and characterization from alkaliphilic *Bacillus licheniformis* KBDL4: A Lonar Soda Lake Isolate. *Indian J. Expt. Biol.*, 2012; 50(8):569-576.
- Khairnar MG, Mahabole MP and Pathak AP. Nanoactivator mediated modifications in thermo stable amylase from *Bacillus licheniformis*. *Ind. J. Biochem and Biophysics*, 2012; 49(6):468-471.
- Deshmukh KB, Taur SA, Cherekar MN, Kothari MN and Pathak AP. Process optimization, purification and characterization of glucoamylase from different Sorghum varieties. *Journal of Chemical and Pharmaceutical Research*, 2011; 3(2):732-737.
- Rathod MG, Pathak AP. Wealth from Waste: Optimized Alkaline Protease Production Using Agro-Industrial Residues by *Bacillus alcalophilus* LW8 and its Biotechnological Applications. *J. Tai bah Univ. Sci.* 2014; 8(4):307-314
- Pathak AP, Rathod MG. Cultivable bacterial diversity of terrestrial thermal spring of Unkeshwar, India. *J. Biochem. Tech.* 2014; 5(4):814-818.
- Pathak AP, Sardar AG. Isolation and characterization of salt stable protease producing archaea from marine solar saltern of

- Mulund, Mumbai. Indian J. Mar. Sci.2014; 43(3):412–417.
12. Sardar AG, Pathak AP. Exploring Microbiota of solar saltern of Mulund, Mumbai, India. Indian J. Mar. Sci. 2012; 43(4):634-641.
  13. Deshmukh MG, Pathak AP. Karuppayil MS. Bacterial diversity of Lonar Soda Lake of India. Indian J Microbiology, 2011; 51(1): 107 – 111
  14. Joshi AA, Kanekar PP, Kelkar A.S., Shouche YS, Vani AA, Borgave SB, Sarnaik SS. Cultivable Bacterial Diversity of Alkaline Lonar Lake, India. Microb. Ecol., 2009; 55(2): 162-172.
  15. Tambekar DH, Tambekar SD. Partial characterization and optimization of alkaline protease production of *Bacillus pseudofirmus* from Lonar Lake. Int. J. Adv. Pharm. Biol. Sci, 2012; 2(1):107-115.
  16. Sinha N, Satyanarayana T. Alkaline protease production by thermophilic *Bacillus licheniformis*. Indian J.Microbiol, 1991; 31(4): 425-430.
  17. Rao RS, Deshmukh YD, Borkar PS, Khobragade CN. Production of alkaline protease from *Bacillus subtilis* using rice bran. J. Cell and Tissue Res, 2008; 8(2): 1347-1350.
  18. Polkade A, Ramana VV, Joshi AA, Pardeshi L, Shouche YS. *Rufibacter immobilis* sp. nov., a novel strain isolated from high altitude saline Lake. International J. Syst. Evol. Microbiol. (In Press) 2015; 65(6):1779-85.
  19. Pandey Anita, Choudhari Shivaji, Sharma Avinash, Choudhary Vipin Singh, Malviya Mukesh Kumar, Chamoli Swati K. Rinu, Trivedi Pankaj, and Palni Lok Man S. Recovery of *Bacillus* and *Pseudomonas* spp. from the 'Fired Plots' under shifting cultivation in northeast India. Current microbiology, 2011; 62(1):273-280.
  20. Aneja KR. Experiments in microbiology, plant pathology and biotechnology, fourth edition. New international publisher, New Delhi, 2005.
  21. Pathak AP and Sardar AG. Isolation and characterization of carotenoid producing haloarchaea from solar saltern of Mulund, Mumbai, India. Indian journal of natural products & resources, 2012; 3(4):483-488.
  22. Tariq\* AL., Reyaz AL. And Prabakaran JJ. Purification and Characterization of 56 KDa cold active Protease from *Serratia marcescens*. African Journal of Microbiology Research, 2011; 5(32): 5841-5847.
  23. Rathod MG and Pathak AP. Isolation and Identification of Alkaline Protease Producer from Selected Alkaline Habitat. International Journal of Innovative Biological Research, 2014; 3(1):1-6.
  24. Mathew\*CD and Gunathilaka RMS. Production, purification and characterization of a thermostable alkaline serine protease from *Bacillus licheniformis* NMS-1. International journal of biotechnology and molecular biology research, 2015; 6(3):19-27.