

**INTERNATIONAL JOURNAL OF ADVANCES IN
PHARMACY, BIOLOGY AND CHEMISTRY****Research Article****Qualitative and Quantitative Determination of
Artificial Sweetener Saccharin Sodium by
FTIR Spectroscopy****Kafa Khalaf Hammud*, Ryadh Raheem Neema,****Issam Shaker Hamza, Essa Abid Salih.**

Ministry of Science and Technology, Baghdad, Iraq

ABSTRACT

Qualitative and quantitative determination of chemicals especially that deal with drugs and pharmaceuticals can be considered as important issues in scientific research to insure human safety and security.

And with this direction of scientific research, saccharin as sodium salt had been determined qualitatively and quantitatively through applying FTIR technique. By applying a weight range of the pure compound, the obtained results showed several absorption bands with standard curves that obeyed Lambert-Beer Law. Also, HPLC technique applied and compared with FTIR of different pharmaceuticals in local Iraqi market.

The E% and Rec. % results for both methods proved that sampling procedure in FTIR technique for both pure and pharmaceutical samples was more superiority beside simplicity.

Keywords: Saccharin, FTIR, HPLC, sweeteners, quantitative determination.

INTRODUCTION

Sweeteners counted as cheap food additives that several times duplicated sugar taste compared with natural sugar (sucrose) with other benefits such as decreasing the resulted energy from food consumption that afforded to weight loss (diet), solving sugar or insulin regulation problems in Diabetes mellitus and Hyperglycemia respectively, and teeth protection¹⁻⁵.

Sweeteners can be classified according to their source to Natural sugar substitutes (natural sweeteners) [Brazzein, Curculin, Mabinlin, Miraculin, Monellin, Pentadin, Thaumatin, Erthritol, Glycyrrhizin, Glycerol, hydrogenated starch hydrolysates, Inulinetc.] (Figure 1) obtained from plants such as proteins, polyols, monosaccharide, extracts with sweetness by weight ranged from 0.4 to 3000¹⁻⁵.

The other classification of sweeteners is Artificial sweeteners or sugar substitutes [Alitame, Aspartame, Dulcin, Glucin, Neotame, P-4000, Saccharin sodium, etc.] (Figure 2) with sweetness by weight range

(30-8000) as a free or K⁺, Na⁺, or Ca²⁺ salt prepared with different methods according to its chemical structure.

Saccharin as sodium salt: 2 – Sodio - 1, 2 – benzisothiazol - 3(2H) - one 1, 1 - dioxide (Figure 3) is artificial sweeteners⁶ applied in several food industries (Teeth pastes, soft drink, candies, and others) and in production of several pharmaceuticals as low calories sweet material. Saccharin sweetness estimated from 300 to 500 times than sucrose and its known as a crystalline white powder with different water contain, soluble in water, slightly soluble in alcohol with different scientific names.

Chem. Abstr. Name: 1,2-Benzisothiazol-3(2H)-one, 1,1-dioxide, sodium salt

IUPAC Systematic Name: 1,2-Benzisothiazolin-3-one, 1,1-dioxide, sodium salt

Synonyms: *ortho*-Benzoylsulfimide sodium salt; saccharin sodium; saccharin sodium salt; saccharin soluble; sodium *ortho*-benzosulfimide; sodium

saccharide; sodium saccharinate; sodium saccharine; soluble saccharin

Saccharin can be prepared as heterocycles according to Remsen-Fahlberg synthesis⁷ (Scheme -1-) or Maumee synthesis¹ (Scheme -2-).

Saccharin does not digested in human and does not help in insulin production as a result of taste⁸. Also, World Health Organization (WHO) through *International Agency for Research on Cancer* (IACR) classified it as Group 2B, possibly carcinogenic to humans then changed it to Group 3, not classifiable as to carcinogenicity to humans. This WHO classification was documented in spite of numerous scientific research pointed to saccharin carcinogenicity on rats, mice, monkeys (especially balder cancer in rats) because WHO considered this material as non-DNA-reactive mechanism⁶.

Despite of the negative effect produced as a result of long term consumption, increasing of world usage and production had been distributed between human personal care, industrial or pharmaceutical products. This matter cleared international scientific research institutes ways to document the suitable analytical methods to determine saccharin sodium such as classical (titration), colorimetric, spectroscopic, electrical, chromatographic methods⁹⁻²¹.

The aim of this work was directed to use FTIR technique for qualitative and quantitative determination of saccharin sodium.

EXPERIMENTAL

Chemicals

All used chemicals were from BDH and were used without further purification. Tablets samples were from local Iraqi market with WHO note of allowed sodium cyclamate (0-12)mg/Kg human weight and sodium saccharin (2.5-5)mg/Kg.

- Sample (A): Dulcaryl from BİLİM PHARMACEUTICALS containing Sodium cyclamate (31.250)mg, Sodium saccharin 2H₂O (3.125)mg
- Sample (B): Sweetcell from MEDCELLPHARMA containing Sweetener: sodium cyclamate, sodium saccharin, lactose

Acidity regulators: sodium hydrogen carbonate, monosodium citrate where sweetness equal to 4.4 gm of sugar.

For standard curve with FTIR technique, (0.4-2.5) mg of standard saccharin sodium was applied. Ten tablets from commercial sweetener were weighted and grinded then calculated for one tablet as an average weight. A weight of the resulted powder equaled to 0.100 mg of pure saccharin sodium was mixed with pure KBr powder.

Instrument

FTIR spectra were recorded using KBr discs on Shimadzu (Japan) IRPestige-21 / FTIR-8000 Fourier Transform Infrared spectrophotometer.

HPLC method was achieved with CeCell instrument with below analytical conditions:

Mobile phase: 50:50 water: methanol with 2 drops of trifluoroacetic acid (TFA).

Column: C18, 25 cm, 4.6 mm i.d x 5 μ L, particle size 3 μ m.

Temperature: room temperature.

Flow rate: 0.7 mL / min.

Detector: U.V at 220 nm

Volume of Injection: 50 μ L

RESULTS AND DISCUSSION

FTIR spectrum of pure saccharin sodium showed several absorption bands coincided with literatures²²⁻²⁷ such as C=O absorption at 1643 cm⁻¹, C-C benzene ring stretching at (1585, 1458) cm⁻¹, -SO₂-N-stretching at (1336, 1257, 1149) cm⁻¹ including asymmetrical and symmetrical -SO₂- stretching vibrations. Negative Saccharinate ion with asymmetrical absorption and carbonyl bending were appeared at (968 and 748) cm⁻¹ respectively.

The resulted FTIR spectra (Figure 4) of accurate weighted pure saccharin sodium gave several standard equations (Table 1) according to different wave numbers with identical R² results. Table 2 showed the efficiency of the applied FTIR method for quantitative determination of saccharin sodium in commercial samples (Figure 5).

The choosing of the proper wave number was based on its representation of special group in saccharin sodium structure did not repeat in other components presented in commercial tablet beside its symmetry with increasing weight with high R² value. On the previous bases, 1643 cm⁻¹ had been selected.

Many literatures²⁸⁻³³ came out HPLC technique applications in qualitative and/ or quantitative determination of saccharin sodium in many different samples and analytical conditions. In this work, HPLC technique had been applied as reference method to determine saccharin sodium where the obtained resulted with range (0.8-4) ppm of pure saccharin sodium gave good standard curve results (R²=0.995) (Figures-6- and -7-) had been used for comparison with FTIR technique results. In HPLC technique, one commercial tablet dissolved in one litre of deionized water (Figure 8).

Tabulated E% and Rec.% results (Table 2 and Table 3) showed that the Turkish sample gave approach values in both applied methods and FTIR method was outdoing than HPLC method in approaching to the recorded values by the manufacture. The final result

for this work with FTIR technique proved that this new quantitative saccharin sodium determination method is with high efficiency compared with the classic HPLC or other analytical method.

CONCLUSIONS

FTIR technique provided with its simplicity in sampling procedure for both pure and commercial samples compared with others. FTIR simplicity and accuracy were obvious in our obtained results in determination saccharin sodium.

Table 1
Standard curve results of each wave number values (cm⁻¹).

Wave number, cm ⁻¹	Standard curve equation	R ²
1643	$y = 1.177x + 0.988$	0.973
1585	$y = 0.137x + 0.075$	0.985
1458	$y = 0.084x + 0.099$	0.992
1336	$y = 0.133x + 0.065$	0.993
1149	$y = 0.280x - 0.018$	0.953
968	$y = 0.184x + 0.033$	0.975
748	$y = 0.170x + 0.024$	0.975

Table 2
Results of applied FTIR method.

Sample name	Tablet weight, mg		FTIR results at 1643 cm ⁻¹		
	Average of one tablet	Applied	Resulted sodium saccharin, mg	E, %	Rec., %
A	42.6	1.125	3.15716	1.029117	101.0291
B	55.9	1.5	2.581398	-17.3953	82.60475

Table 3
Results of applied HPLC method.

Sample name	One tablet weight, mg	HPLC results at 4.17 min. as a retention time		
		Resulted sodium saccharin, mg	E, %	Rec., %
A	42.6	4.58907	46.85023	146.8502
B	55.9	6.739814	115.674	215.674

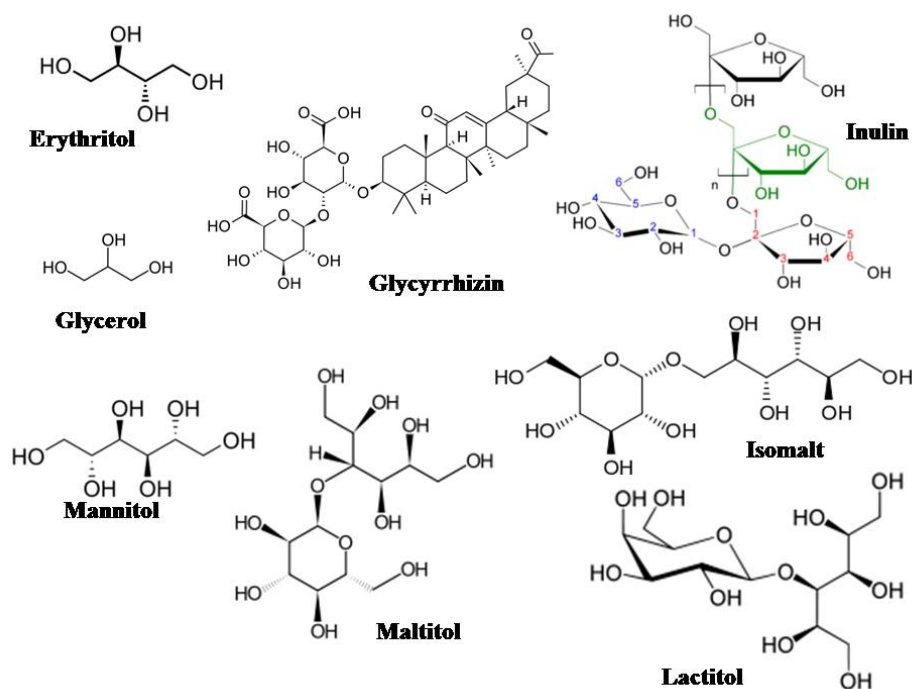


Figure 1
Chemical structures of several natural sweeteners.

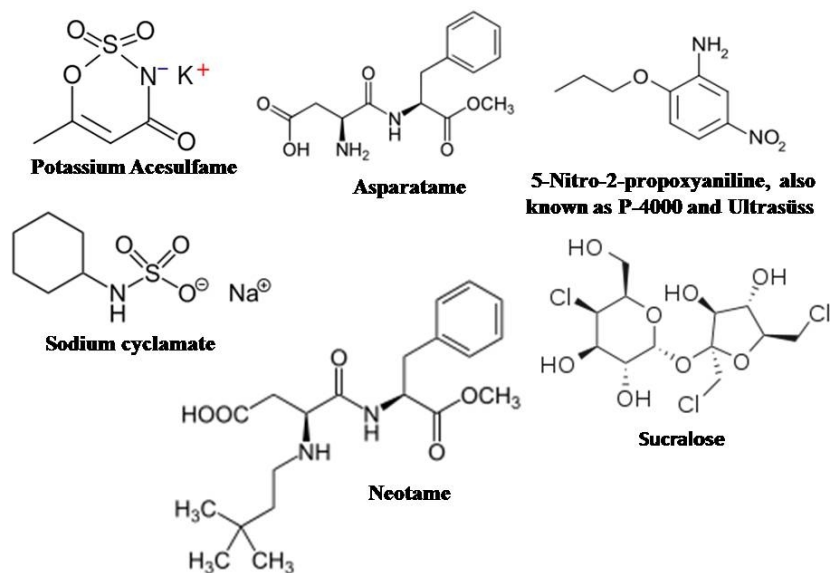


Figure 2
Chemical structures of several artificial sweeteners or sugar substitutes.

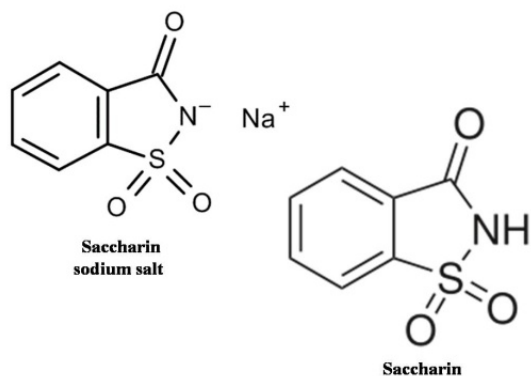
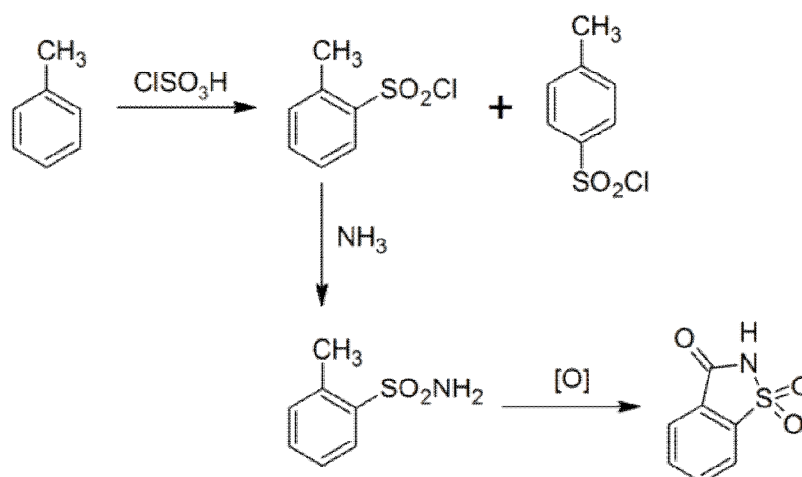
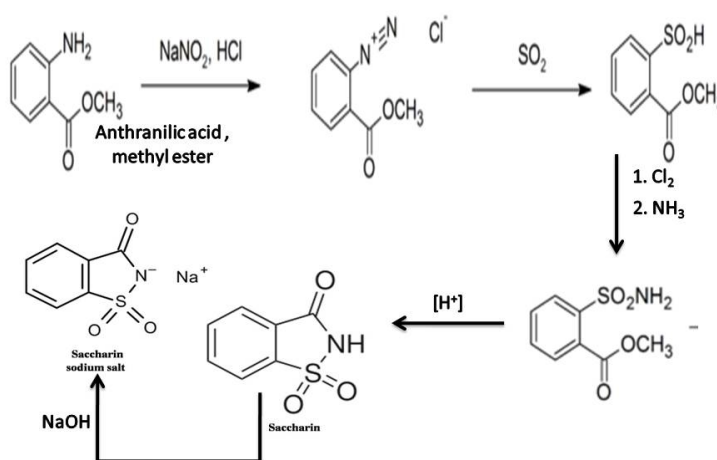


Figure 3
Saccharin and its salts chemical structures.



Scheme 1
Remsen-Fahlberg synthesis.



Scheme 2
Maumee synthesis.

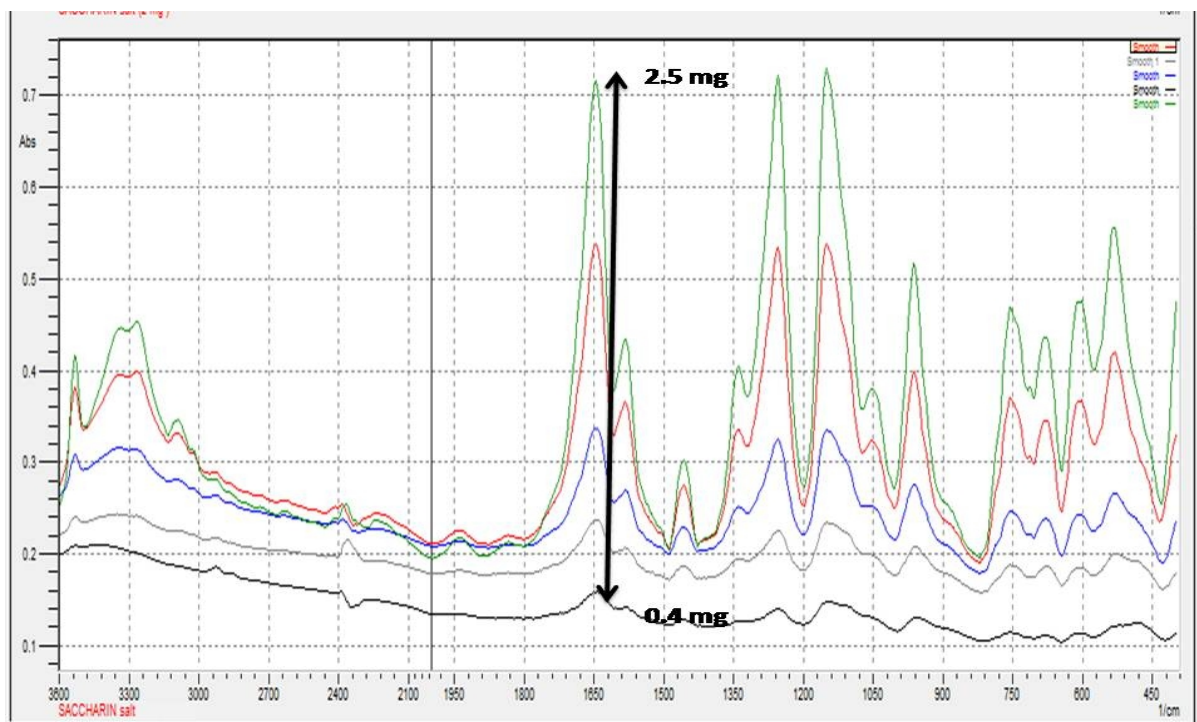


Figure 4
FTIR spectra for saccharin sodium with different weights (0.4-2.5 mg).

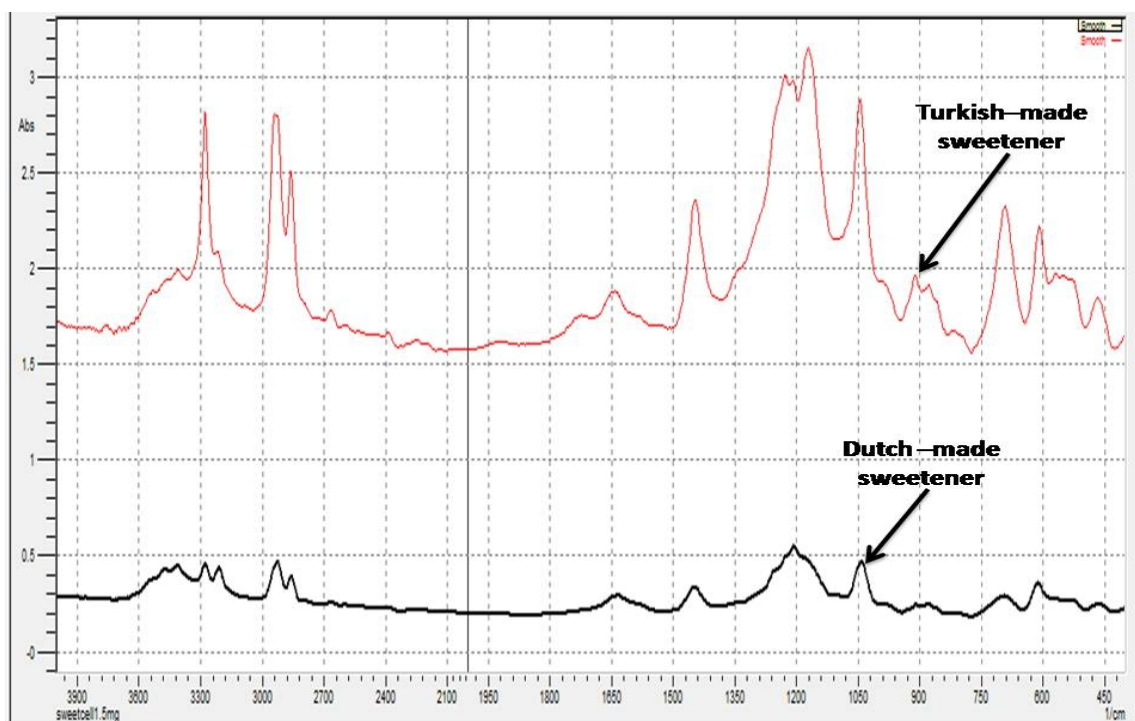


Figure 5
FTIR spectral for two commercial sweeteners containing saccharin sodium.

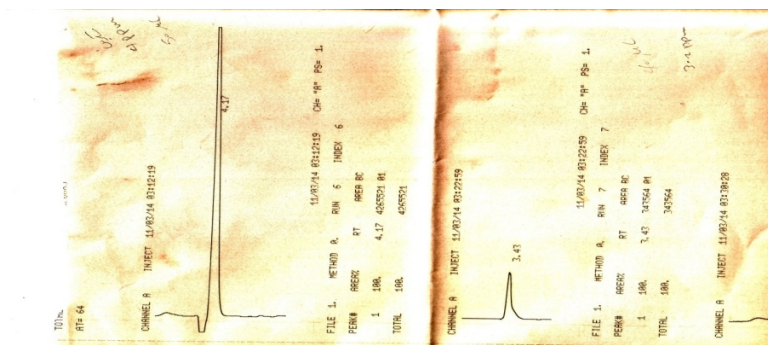


Figure 6
HPLC chromatogram for pure saccharin sodium (3.2 and 4) ppm

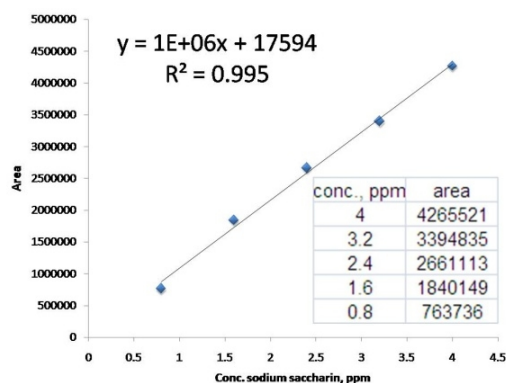


Figure 7
HPLC standard curve for saccharin sodium.

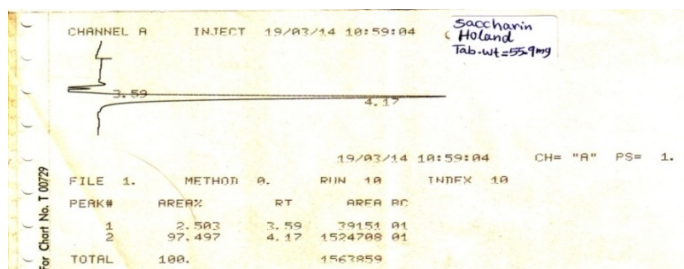
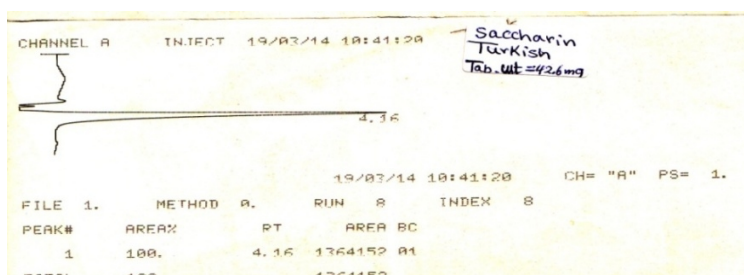


Figure 8
HPLC chromatogram for two commercial sweeteners (Turkish-made (A) and Dutch-made (B)) samples

REFERENCES

1. Lipinsk, G., 2005. Sweeteners: Ullmann 's Encyclopedia of Industrial Chemistry, Wiley-VCH, Weinheim:, Germany.
2. Jang H, Kokrashvili Z, Theodorakis M., Carlson O, Kim B, Zhou J, Kim H, Xu X, Chan S, Juhaszova M, Bernier M, Mosinger B, Margolskee R, Egan J, Gut-expressed gustducin and taste receptors regulate secretion of glucagon-like peptide-1, Proc. Natl. Acad. Sci. (USA) 2007; 104 (38): 15069-15074.
3. Frank G, Oberndorfer T, Simmons A, Paulus M, Fudge J, Yang T, Kaye W, Sucrose activates human taste pathways differently from artificial sweetener, Neuroimage 2008; 39: 1559-1569.
4. Mezitis N, Maggio C, Koch P, Quddoos A, Allison D, Pi-Sunyer F, Glycemic effect of a single high oral dose of the novel sweetener sucralose in patients with diabetes, Diabetes Care 1996; 19(9): 1004-1005.
5. Ma H, Bellon M, Wishart J, Young R, Blackshaw L, Jones K, Horowitz M, Rayner C, Effect of the artificial sweetener, sucralose, on gastric emptying and incretin hormone release in healthy subjects, AJP: Gastrointest. Liver Physiol. 2009; 296 (4): G735-G739.
6. World Health Organization (WHO) - International Agency for Research on Cancer (IARC) Monographs, 1999. Some chemicals that cause tumours of the kidney or urinary bladder in rodents and some other substances. Volume 73, , Lyon, France: WHO Press, p.517-624.
7. Ager D, Pantaleone D, Henderson S, Katritzky A, Prakash I, Walters D, Commercial, Synthetic Nonnutritive Sweeteners, Angew. Chem. Int. Ed. 1998; 37 (13-24): 1802-1817.
8. Ionescu E, Rohner-Jeanrenaud F, Proietto J, Rivest R, Jeanrenaud B, Taste-induced changes in plasma insulin and glucose turnover in lean and genetically obese rats, Diabetes 1988; 37(6): 773-779.
9. Mitchell L, Separation and identification of cyclohexylsulfamate, dulcin, and saccharin by paper chromatography, J. Assoc. Off. Agric. Chem. (Washington) 1955; 38: 943-947.
10. Neiman M, Polarographic determination of saccharin, J. Anal. Chem. USSR Engl. Transl. 1955; 10: 163-167 (1955).
11. Markus J, Gravimetric determination of saccharin in cider, J. Assoc. Off. Anal. Chem. 1973; 56:162-163.
12. Fix G., Pollack J, Determination of saccharin in watts nickel plating solutions by first derivative ultraviolet spectrometry, Anal. Chem. 1980; 52: 1589-1592.
13. Riggan R, Kinzer G, Characterization of impurities in commercial lots of sodium saccharin produced by Sherwin-Williams process. I. Chemistry, Food Chem. Toxicol. 1983; 21: 11-17.
14. Ramappa P, Nayak A, Rapid spectrophotometric determination of saccharin in soft drinks and pharmaceuticals using azure B as reagent, Analyst 1983; 108: 966-970.
15. Guven K, Ozol T, Ekiz N, Guneri T, Spectrophotometric determination of sodium cyclamate and saccharin sodium with astrazone pink FG, Analyst, London 1984; 109: 969-970.
16. Hernandez-Cordoba M, Lopez-Garcia I, Sanches-Pedreno C, Spectrophotometric determination of saccharin in different materials by a solvent extraction method using Nile blue as reagent, Talanta, London 1985; 32: 325-327.
17. J. Hann J, Gilkison I, Gradient liquid chromatographic method for the simultaneous determination of sweeteners, preservatives and colours in soft drinks, J. Chromatogr. 1987; 395: 317-322.
18. Auricchio M, Batistic M, Hoppen V., Yamashita I, Pesquisa de adoçantes não calóricos sintéticos em adoçante natural de *Stevia rebaudiana* (Bert) Bertoni, Rev. Bras. Farmacogn., São Paulo 1989; 2-4: 53-61.
19. Schapoval E, Controle biológico de qualidade de produtos farmacêuticos, correlatos e cosméticos, Rev. Bras. Cienc. Farm. 2005; 41(2): 279-280.
20. US Pharmacopeia. 25th ed. Rockville: United States Pharmacopeial Convention, p.1545, 2610 (2002).
21. British pharmacopeia London: Her Majesty's Stationery Office, p.495-496 (1988).
22. Baxter J, Craig J, Willis J, The infrared spectra of some sulphonamides, J. Chem. Soc. 1955; 669-679.
23. O'Sullivan D, Vibrational frequency correlations in heterocyclic molecules. Part VI. Spectral features of a range of compounds possessing a benzene ring fused to a five-membered ring, J. Chem. Soc. 1960; 3278-3284.
24. Nabar M., Khosla A, Studies on saccharinates I. Synthesis and some properties of rare earth saccharinate hydrates, J. Alloy. Compd. 1995; 225: 377-380.
25. Yueng Z, Spectral and thermal properties of saccharin and saccharin metal complexes, Transit. Met. Chem. 1994; 19(4): 446-448.

26. Naumov P, Jovanoski G, Vibrational Study of two Novel Cesium Saccharinates. Spectroscopic Evidence for Organic Molecule Embedded in Ionic Salt, *Vibr. Spectrosc.* 2000; 24(2): 201-211.
27. Çakır S, Odabaşoğlu M, Biçer E, Yazar Z, Synthesis, spectroscopic and voltammetric studies of a novel Schiff-base of cysteine and saccharin, *J. Mol. Struct.* 2009; 918: 81-87.
28. Wasik A, McCourt J, Buchgraber M, Simultaneous determination of nine intense sweeteners in foodstuffs by high performance liquid chromatography and evaporative light scattering detection—Development and single-laboratory validation, *J. Chromatogr. A* 2007; 1157(1-2): 187-196.
29. Ferrer I, Thurman M, Analysis of sucralose and other sweeteners in water and beverage samples by liquid chromatography/time-of-flight mass spectrometry, *J. Chromatogr. A* 2010; 1217(25): 4124-4134.
30. George V, Arora S, Wadhwa B, Singh A, Analysis of multiple sweeteners and their degradation products in lassi by HPLC and HPTLC plates, *J. Food Sci. Technol.* 2010; 47(4): 408-413.
31. Mahboubifar M, Sobhani Z, Dehghanzadeh G, Javidnia K, A Comparison between UV Spectrophotometer and High-performance Liquid Chromatography Method for the Analysis of Sodium Benzoate and Potassium Sorbate in Food Products, *Food anal. Method* 2011; 2: 150-154.
32. Zyglar A, Wasik A, Wasik A, J. Namieśnik, Determination of nine high-intensity sweeteners in various foods by high-performance liquid chromatography with mass spectrometric detection, *Anal. Bioanal. Chem.* 2011; 400(7): 2159-2172.
33. Grembecka M, Szefer P, Simultaneous Determination of Caffeine and Aspartame in Diet Supplements and Non-Alcoholic Beverages Using Liquid-Chromatography Coupled to Corona CAD and UV-DAD Detectors, *Food anal. Method* 2012; 5: 1010-1017.